

## STUDIES OF LEAD RESIDUES IN BONES AND FEATHERS OF GRIFFON VULTURES

Petar Stamberov, Tandzhu Mehmedov

*University of Forestry, Faculty of Veterinary Medicine, Sofia, Bulgaria*

*E-mail: [pstamberov@ltu.bg](mailto:pstamberov@ltu.bg)*

### ABSTRACT

The aim of the present research is to study the content of lead in feathers and bone samples in eight carcasses of Griffon Vultures (*Gyps fulvus*, Habitz 1870) from a region in southwestern Bulgaria. In order to exclude surface lead contamination, the feather samples were preliminary examined by X-ray fluorescent spectroscopy. The samples of lead concentration in feathers and bones were analyzed by the methods of ICP-MS Spectrometry. The results displayed residues of lead in the feathers and bones only in some individuals.

**Key words:** Griffon vultures, lead, samples, bones, feathers, ICP-MS Spectrometry.

### Introduction

Incidents of lead poisoning in birds have been reported from all around the world (Fisher et al. 2006; Bounas et al. 2016). Birds of prey are one of the main groups affected by exposure to lead impacting several threatened species (Mateo, 2009). Like other vultures, the Griffon vulture is a scavenger, that consume died game species may ingest lead fragments or shot embedded in their prey's flesh (Mateo et al. 1997; Pain et al. 1997).

The Griffon Vulture (*Gyps fulvus*, Habitz 1870) has a large breeding range, extending over Europe, the Middle East and North Africa. A reintroduction Vultures Return in Bulgaria LIFE08NATY/BG/278 Project is underway since 2010 in Stara Planina Mountain, Bulgaria (Deinet et al. 2013; Botha et al. 2017).

The feathers must be used with care because the complex, fine structure is highly prone to accumulating surface contamination (Burger, 1993; Dauwe et al., 2003; Golden et al., 2011; Jenny et al., 2015). Lead accumulates in feathers through active or passive diffusion from the blood into the feather follicle (Cardiel et al., 2011).

Lead in bone has been physiologically incorporated over the lifetime of the bird (Franson and Pain, 2011). Following absorption, lead is deposited in a range of tissues, primarily liver, kidney and bone. Whilst lead in liver and kidney has a turnover rate of weeks to months, lead is retained in bone for years, thus reflecting both acute and lifetime chronic exposure from all sources (Garcia-Fermindez et al. 1997; Pain et al. 2005).

In this article, we analyzed the concentrations of lead in the bones and feathers of the griffon vultures in order to determine residues of lead and evaluate the extent to which this species may be subject to lead poisoning.

### Materials and methods

The feathers and bone samples in eight carcasses of Griffon Vultures from a region in southwestern Bulgaria in cooperation with the Wildlife Rescue Center "Green Balkans" in Stara Zagora were obtained. The cause of death was illegal poisoning with carbamate pesticides.

The collected feathers were examined by X-ray fluorescence spectrometry (XRF) for detection of exogenous incorporated lead. X-ray diffraction spectrometer *Fischerscope*® with high-resolution

CCD color camera for optical monitoring of the measurement location along the primary beam axis were used. Then feathers were washed in running deionized water and air dried. Then the feathers were cutting and isolated in airtight plastic bags and refrigerated at 0–4 °C for later analysis.

Bone samples of approximately 3–5 g were extracted from the femur of each of the vultures. The bones were removed from the muscles and bone marrow with scalpel and bone scissors. Then the bones were dried to a constant weight (0.5–1 g) and stored in sterile containers and refrigerated at -20 °C until the lead analysis.

The survey was conducted in the Laboratory of Ecology and technical tests "Aquaterratest" Sofia. The samples were analyzed by the methods of inductively coupled plasma spectrometry. The spectrometers Varian Vista-MPX CCD Simultaneous ICP OES, Varian Australia, and Plasma Quant MS S-NR 105000-AQ032, *Analytik Jena AG*, Germany were used. The detection limit was 0.1 mg/kg dry-weight.

## Results

The preliminary X-ray fluorescence spectrometry (XRF) excludes the presence of exogenous incorporated lead in none of the feather samples were examined (Fig. 1).

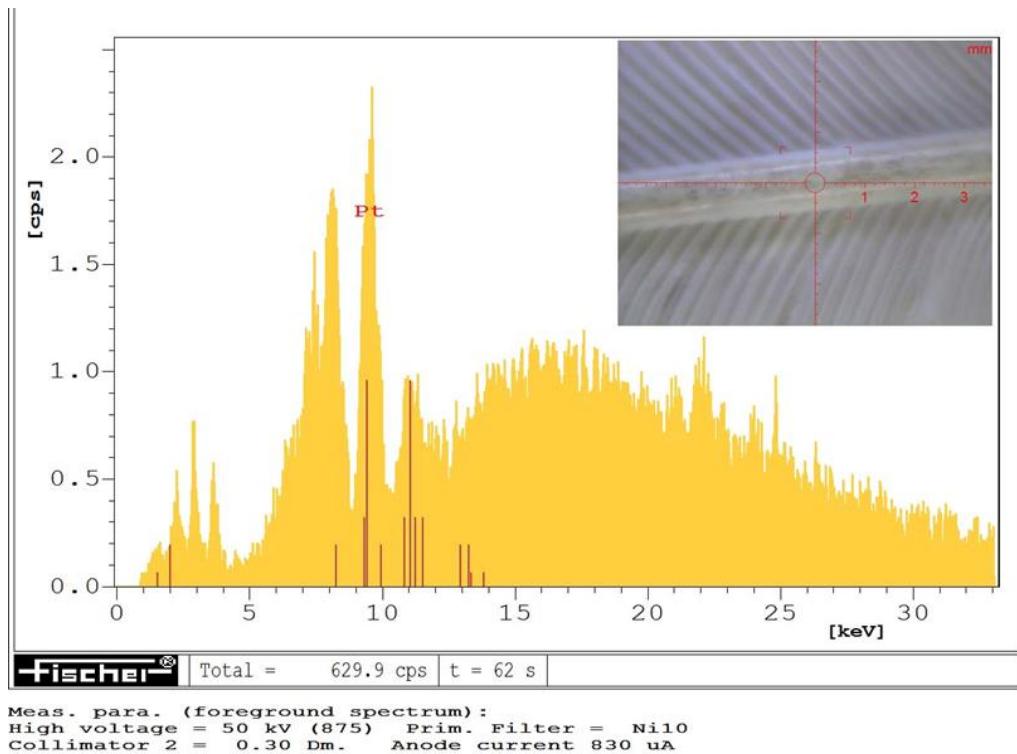


Figure 1: XRF of surface the feather of Griffon Vultures does not show presence of lead. The presence of platinum in some samples cannot be explained. The vaguely defined peaks after 15 keV are result to the large amount of light elements such as Ca, P and Mg. The presented figure is indicative for a series of measurements of different samples.

### Lead concentration in feathers

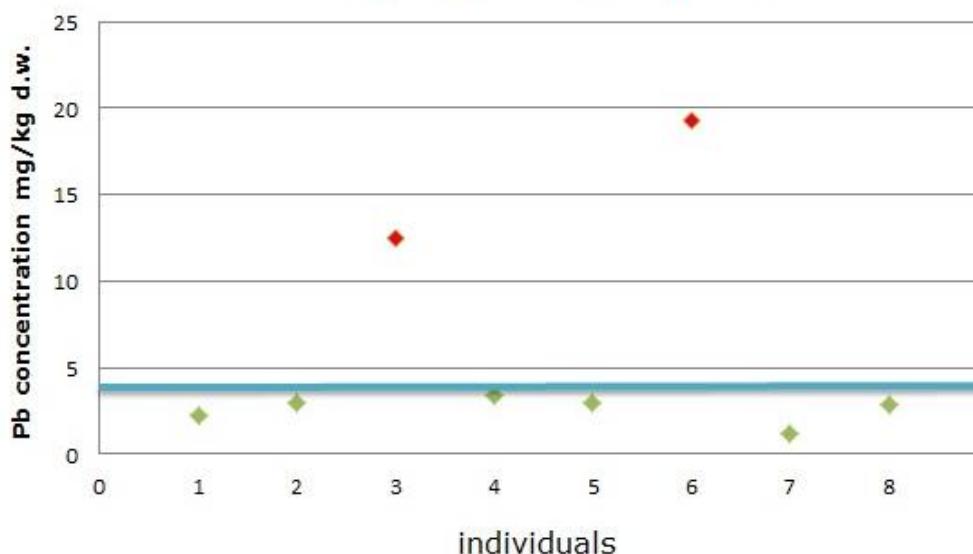


Figure 2: Concentrations of lead in feathers of griffon vultures (*Gyps fulvus*, Habitz 1870). Tolerable threshold of lead content in feathers is marked with solid blue line (4 mg/kg d.w.). The high lead concentration samples (12.49 and 19.81 mg/kg d.w.) are marked with red color.

### Lead concentrations in bones

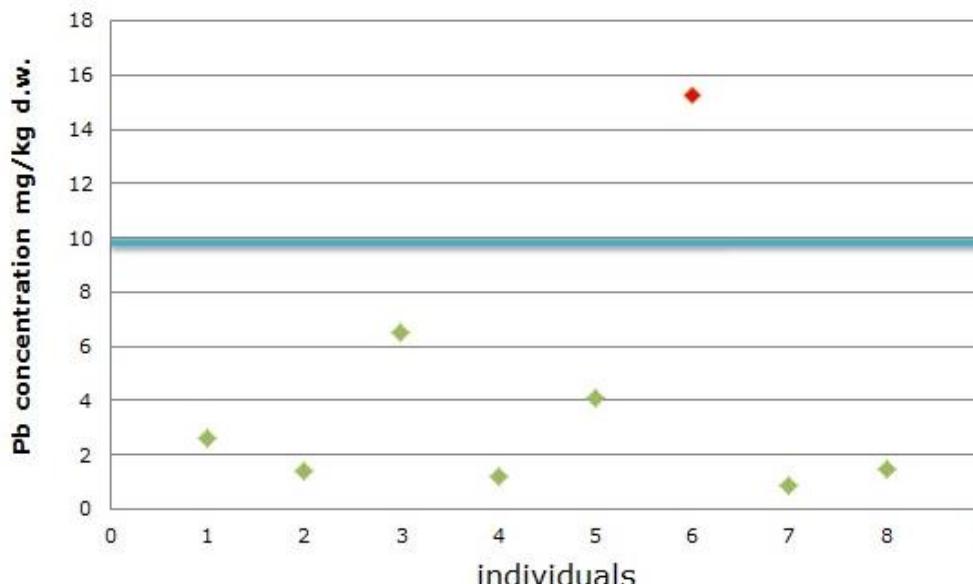


Figure 3: Concentrations of lead in femoral bones of griffon vultures (*Gyps fulvus*, Habitz 1870). Tolerable threshold of lead content in bones (<10 mg/kg d.w.) is marked with solid blue line. The sample with high lead levels is marked with red color.

Two (No 3 and No 6) of the eight feather samples showed high lead concentration of 12.49 and 19.81 mg/kg d. w. according to tolerable risk threshold of 4 mg/kg d. w. In four samples (No. 2, 4, 5 and 8) the feathers lead concentration showed about and slightly below threshold levels of 4 mg /kg d. w. The other two samples (No. 1 and 7) showed lead concentration in the feathers low of threshold levels of 4 mg /kg d. w. (Fig. 2).

The analysis of lead content in the femoral bones showed that in seven of the eighth samples the lead levels were below the background concentrations (<10.00 mg/kg d.w). A high lead concentration of 15.21 mg/kg d.w. was registered in only one (No. 6) corresponding to the high lead concentration of the same sample (No 6) in the feather. Sample No. 3 also showed some elevated lead content in bones corresponding to the high concentration of lead in the feathers of the same sample (Fig. 3).

## Discussion

The distribution of lead concentrations in feathers is variable. The results should have been carefully interpreted due to the fact that, like the coat of mammals, the feathers of birds are exposed to anthropogenic pollution (Burger, 1993; Pain et al., 2005). X-ray fluorescence spectrometry has showed absence of surface lead contamination in feathers. Threshold value of > 4 mg/kg of lead concentrations in feathers is associated with adverse effects (Cardiel et al., 2011). According to Jenny et al. (2015), such presence of lead in the feathers may be a result of possible cumulative effects of lead due to repeated oral exposure.

Bone lead concentration is generally considered the best indicator of lead exposure over the total lifetime of the birds (Franson and Pain, 2011). This concentration is more difficult to interpret and threshold the value could be higher because of accumulation (Szymczak and Adrian, 1978). Lead in bones is relative immobile and excretion is very slow (Scheuhammer, 1987). The researchers recommend that lead concentrations (dry weight basis) of <10 mg/kg in bone may be considered background, and >10 mg/kg may indicate elevated lead exposure (above background levels). Birds with bone lead concentration of  $\leq$  20 mg/kg d.w be considered evidence of subclinical to clinical poisoning, and >20 mg/kg d.w be considered evidence of severe clinical poisoning (Mateo et al., 2003; Martin et al. 2008; Franson and Pain, 2011). Accordingly, in our case a high lead concentration in bones of one individual may be indicated for elevated lead exposure.

## Conclusion

The results of this research displayed residues of lead in the feathers and bones only in some individuals. However, the feathers and bones provide a useful screening biomonitoring and potentially can be useful for future studies.

The establishment of facilities that can perform toxicological analysis of vultures is needed in our country in order to assess the threat, since any mortality that can be avoided, especially in such a small population.

## Acknowledgments

This research work was carried out with support of University of Forestry and project BG05M2OP001-2.009-0034 "Support for the development of scientific capacity in the University of

Forestry“, Operational Program "Science and Education for Smart Growth", cofunded by the European Union through the European Structural and Investment Funds.

We also would like to express our gratitude to the colleagues from Green Balkans Rescue Center, Stara Zagora for the provided samples.

## References

1. Botha, A. J., Andevski, J., Bowden, C. G. R., Gudka, M., Safford, R. J., Tavares, J. and Williams, N. P. (2017). *Multi-species Action Plan to Conserve African-Eurasian Vultures*. CMS Raptors MOU Technical Publication No. 5. CMS Technical Series No. xx. Coordinating Unit of the CMS Raptors MOU, Abu Dhabi, United Arab Emirates.
2. Bounas, A., Ganoti, M., Giannakaki, E., Akrivos, A., Vaylyis, D., Zorrilla, I., Saravia, V. (2016). *First confirmed case of lead poisoning in the endangered Egyptian Vulture (Neophron percnopterus) in the Balkans*. Vulture News, 70: 22–29.
3. Burger, J. (1993). *Metals in avian feathers: Bioindicators of environmental pollution*. Reviews of Environmental Contamination and Toxicology, 5, 203–311.
4. Cardiel, I. E., M. A. Taggart, R. Mateo, (2011). *Using Pb-Al ratios to discriminate between internal and external deposition of Pb in feathers*. Ecotoxicology Environmenal Safety, 74, 911–917.
5. Dauwe, T., Bervoets, L., Pinxten, R., Blust, R., Eens, M., (2003). *Variation in heavy metals within and among feathers of birds of prey: effects of molt and external contamination*. Environmental Pollution 124, 429–436.
6. Deinet, S., Ieronymidou, C., McRae, L., Bur eld, I. J., Foppen, R. P., Collen, B. and Buhm, M. (2013). *Wildlife comeback in Europe: The recovery of selected mammal and bird species. Final report to Rewilding Europe by ZSL*. BirdLife International and the European Bird Census Council. Zoological Society of London, London.
7. Golden, N. H., B. A. Rattner, J. B. Cohen, D. J. Hoffman, E. Russek-Cohen, M. A. Ottlinger. (2003). *Lead accumulation in feathers of nestling black-crowned night herons (Nycticorax nycticorax) experimentally treated in the field*. Environmental Toxicology and Chemistry, 22, 1517–1524.
8. Fisher, I. J., D. J. Pain, V. G. Thomas. (2006). *A review of lead poisoning from ammunition sources in terrestrial birds*. Biological Conservation 131, 3, 421–432.
9. Franson J. C., D. J. Pain. (2011). *Lead in birds*. In: Beyer W. N., Meador J. P. (eds). Environmental contaminants in biota: interpreting tissue concentrations. Taylor & Francis Group, Boca Raton, Florida, pp. 563–593.
10. García-Fernández, A. J., M. Motas-Guzman, I. Navas, P. Maria-Mojica, A. Luna, J. A. Sanchez-Garcia. (1997). *Environmental exposure and distribution of lead in four species of raptors in south-eastern Spain*. Archives Environmental Contamination Toxicology, 33:76–82.
11. Jenni, L., M M. Madry, T. Kraemer, J. Kupper, H. Naegeli, H. Jenni, D. Jenni. (2015). *The frequency distribution of lead concentration in feathers, blood, bone, kidney and liver of golden eagles Aquila chrysaetos: insights into the modes of uptake*. Journal of Ornithology, Published online, 14 April 2015. Doi: 10.1007/s10336-015-1220-7.
12. Martin, P. A., D. Campbell, K. Hughes, and T. McDaniel. (2008). *Lead in the tissues of terrestrial raptors in southern Ontario, Canada, 1995–2001*. Science of The Total Environment, 391 (1), 96–103.
13. Mateo, R., R. Molina, J. Grifols, R. Guitart. (1997a). *Lead Poisoning in a Free Ranking Griffon Culture (Gyps fulvus)*. Veterinary Record, 140, 47–48.
14. Mateo, R., M. Taggart, A. A. Meharg. (2003). *Lead and Arsenic in Bones of Birds of Prey from Spain*. Environmental Pollution, 126, 107–114.
15. Mateo, R. (2009). *Lead poisoning in wild birds in Europe and the regulations adopted by different countries*. In: R. T. Watson, M. Fuller, M. Pokras, and W. G. Hunt (Eds.). *Ingestion of Lead from Spent Ammunition, Implications for Wildlife and Humans*. The Peregrine Fund, Boise, Idaho, USA.

DOI 10.4080/ilsa.2009.0107.

16. Pain, D. J., C. Bavoux, G. Burneveau. (1997). *Seasonal blood lead concentrations in marsh harriers Circus aeruginosus from Charente-Maritime, France: relationship with the hunting season*. Biology Conservation, 81, 1–7.
17. Pain, D. J., A. A. Meharg, M. Ferrer, M. Taggart, V. Penteriani. (2005). *Lead concentrations in bones and feathers of the globally threatened Spanish imperial eagle*. Biology Conservation, 121, 603–610.
18. Scheuhammer, A. M. (1987). *The chronic toxicity of aluminum, cadmium, mercury, and lead in birds: a review*. Environmental Pollution, 46, 263–295.
19. Szymczak, M. R., W. J. Adrian. (1978). *Lead poisoning in Canada Geese in Southeast Colorado*. Journal of Wildlife Management, 42, 299–306.